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Oxime and Oxime Ether Derivatives of 1,4-Benzothiazine Related to Oxiconazole

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The synthesis, in vitro antifungal activity, and molecular docking experiments of some oxime and oxime ether derivatives of azole 1,4-benzothiazine are reported herein, with the aim of evaluating

the influence of a partially constrained scaffold that is structurally related to Oxiconazole and bearing the 1,4-benzothiazine moiety, on the inhibition of Candida albicans CYP51.

Introduction

The opportunistic human pathogen *Candida albicans* and other non-albicans species have acquired considerable significance in recent years because of the enhanced susceptibility of immunocompromised patients.^[1] These pathogenic species of *Candida* derive their importance

Figure 1. Structures of Oxiconazole, target compounds, and reference ether derivatives.

not only from the severity of their infections but also from their ability to develop resistance against antifungal drugs.^[2] Therefore, the need to develop new antifungal drugs is higher than ever

Recently we reported the synthesis and antifungal activity of a series of *C. albicans* CYP51 inhibitors^[3] related to Econazole and Ketoconazole based on the replacement of the 2,4-dichlor-ophenyl group with the 1,4-benzothiazine moiety that, in itself, shows some antifungal activity.^[4]

Herein, we report the synthesis and in vitro antimycotic activity of some oxime and oxime ether derivatives of azole 1,4-benzothiazine **3a-g** and **4a-g** related to Oxiconazole and to our previously synthesized compounds. The aim was to extend the current structure–activity relationship (SAR) of *C. albicans* CYP51 inhibitors by investigating the effect of partially constraining the molecule through the insertion of oxime and oxime ether groups.

Oxiconazole (Oxistat®),^[5] is a well-known antifungal agent with a broad spectrum of activity, used for the treatment of skin mycoses. It is structurally characterized by an oxime ether group in the Z configuration. Several compounds containing an oxime or an oxime ether function have been reported to exhibit antimicrobial activity.^[6] (see Figure 1)

Initially our attention was focused on the modification of compound **5 a** as it had shown an appreciable inhibitory activity on *C. albicans* growth, a very good in vivo antimycotic activity combined with an interesting capability to stimulate the immune response. [3a,b]

Results and Discussion

The synthesis of compound **4a** was planned and carried out starting from the known ketone **2a**^[3a] that was first converted into the corresponding oxime derivative **3a** by reaction with hydroxylamine hydrochloride and then alkylated with 4-chlorobenzyl chloride (see Scheme 1).

The increased antifungal activity of 4a (MIC = 15.6–7.8 μ g mL⁻¹ against *C. albicans* CA-6) with respect to 5a (MIC = 46 μ g mL⁻¹) motivated us to extend this structural modification to a series of compounds with a different lateral chain position, different substituents at the C2 position of the nucleus-base, a triazole instead of the imidazole, or longer alkyl substituents

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Supporting information for this article is available on the WWW under http://www.chemmedchem.org or from the author. Elemental analysis for target compounds 3a-g, 4a-g and 5d. Synthetic pathway to obtain compound 5d.

Scheme 1. Synthesis of oxime and oxime ether derivatives and corresponding ether compounds: a) NH₂OH·HCl; b) NaH, 4-chlorobenzyl chloride.

on the cyclic amide, thus obtaining compounds 3a-g and 4b-g.

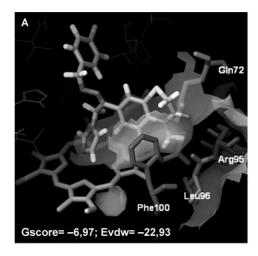
Ketones **2a**–**g** were the key intermediates for the synthesis of the new compounds. Whereas derivatives **2a**–**c** and **2e**–**g** were recently synthesized, Compound **2d** was obtained starting from 7-acetyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one, Collowing a synthetic pathway similar to the one previously reported. Moreover, to be able to have a direct comparison between the activities of ether and oxime ether compounds, the *N*-butyl derivative **5d**, to date unreported, was synthesized from ketone **2d** by reduction with NaBH₄ and subsequent alkylation with 4-chlorobenzyl chloride. The azole ketones **2a**–**g** were converted into the corresponding oxime derivatives **3a**–**g** by reaction with hydroxylamine hydrochloride in pyridine or in EtOH/NaOH. The choice of reaction conditions depended on the differences in solubility between the 6- and 7-substituted 1.4-benzothiazines.

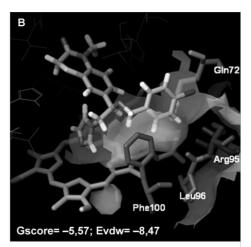
Z/E configurations of the oximes thus obtained were established by comparing their ¹H NMR spectra and the literature data. The 7-substituted 1,4-benzothiazines 3a-d gave the Z isomer as a single product; their ¹H NMR spectra showed only one signal for the methylene protons, and only one signal attributable to the oxime proton; the values of these signals are consistent with the Z configuration. [6a] Moreover, the syn relationship was confirmed by Overhauser correlations measured between the oxime proton and the methylene ones. Compounds **3 e-g** were obtained as *Z/E* mixtures, as deduced from their ¹H NMR spectra. For compound 3 g, for instance, the ¹H NMR spectrum of the mixture showed two signals as singlets at $\delta = 5.10$ and 5.25 ppm (1.4 H and 0.6 H, respectively), corresponding to the protons in position 2'. The most unshielded signal was attributed to the Z isomer, because of the presence of the nearby hydroxyl function. [6e] Similar signals were observed for compounds 3e and 3f. Based on these findings, the configuration of each compound was assigned by repeating ¹H NMR experiments after the separation of the Z/E mixtures. With regard to the Z/E ratio, the E isomer was only predominant for compound 3 g (Z/E ratio 3:7), whereas the Z isomer predominant for both was oximes 3e (Z/E ratio 9:1) and 3f (Z/E ratio 7:3). The high ratio did not allow the E isomer to be isolated for compound 2e. The oximes 3a-g thus obtained and characterized were then converted into the corresponding 4chlorobenzyl ether derivatives 4a-g by reaction with 4-chlorobenzyl chloride in the presence sodium hydride of (see Scheme 1). ¹H NMR spectra of the final products confirmed the retention of the configuration, in accord with the literature data.[8]

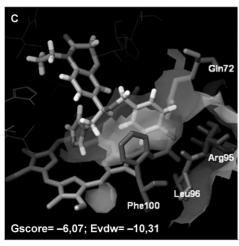
The in vitro antifungal activity against *C. albicans* and *C. krusei* of compounds 3a-g and 4a-g was evaluated in comparison with their corresponding previously synthesized ether derivatives 4a-g. Unfortunately, the microbiological data confirmed a good activity only for the oxime ether derivative (*Z*)-4a.

To gain some insight into the structure-activity relationships and binding mode of compounds 4a, 5a, 4c, and 5c, a number of docking experiments were performed in a 3D model of C. albicans CYP51 as detailed in the experimental section. The results partially clarified the different activities observed among the selected compounds. In particular, the main variation was in the different binding modes adopted by the compounds at the active site of the enzyme. Whereas the access channel to the catalytic site is filled with the 4-chlorophenyl side chain in the case of the N-methyl ether derivative 5a and N-ethyl ether derivative 5c, [9] the N-methyl 1,4-benzothiazine moiety occupies the channel in the binding mode of the oxime ether analogue 4a. This latter orientation seems to favor the binding of compound 4a over 5a and 5c at the enzyme (glide scores for 4a=6.97, 5a=5.57, 5c=6.07). In particular, the contribution made by the steric interaction term (van der Waals energy, Evdw) of compound 4a to the binding energy was about twice that of 5c and three times that of compound 5 a. These results indicate that the molecular shape of this compound has a good fit within the enzymatic binding site (see Figure 2).

Despite the good activity displayed by the ether derivative $\mathbf{5c}$ (MIC = $15.6 \, \mu \mathrm{g \, mL^{-1}}$), the *N*-ethyl oxime ether $\mathbf{4c}$ was inactive. Docking results of $\mathbf{4c}$ indicated that there were no possible binding modes present in the catalytic site of CYP51 for this molecule. The failure of the docking algorithm to identify a binding pose for compound $\mathbf{4c}$ can be ascribed to the presence of two structural features (Figure 2B): 1) the small size of the binding pocket (residues Gln72, Arg 95, Leu 96, and







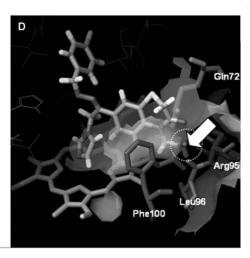


Figure 2. Docking results of studied compounds within the catalitic site of CYP51. The residues lining the pocket of the binding site are in stick style and their van der Waals molecular surface is highlighted. The images A, B, and C show the results of derivatives **4a**, **5a**, and **5c**, respectively. In D, the hypothetical pose of compound **4c** is shown to clarify the steric clashes of the *N*-ethyl group (highlighted with white point circle) in the small pocket constituted by residues Gln72, Arg 95, Leu 96, and Phe 100.

Phe 100) that hosts the *N*-methyl 1,4-benzothiazine moiety of compound **4a** does not allow the enzyme to accept the *N*-ethyl 1,4-benzothiazine group of **4c** and 2) the structural stiffness of the oxime function of the molecule that hampers any conformational fitness within the above described enzymatic pocket.

Conclusions

As a continuation of an articulated program, the aim of this work was twofold: 1) to synthesize new azole derivatives containing a 1,4-benzothiazine nucleus, that, in itself, shows a moderate antifungal activity and 2) to improve the antifungal activity and extend the SAR for this class of compounds.

As a result of structural change, it was possible to establish that the side chain that bears the azole nucleus must be flexible, as its constraint in a more rigid moiety generally proved to be detrimental to the activity.

Experimental Section

Melting points determined in capillary tubes (Electrothermal, model 9100, melting point apparatus) were uncorrected. Elemental analysis was performed on a Carlo Erba element analyzer 1106, and the data for C, H, and N are within $\pm 0.4\%$ of the theoretical values. ¹H NMR spectra were recorded at 200 MHz (Bruker AC-200 spectrometer) or at 400 MHz (Bruker Avance-DRX 400 spectrometer) with Me₄Si as internal standard. Chemical shifts are given in ppm (δ) , and the spectral data are consistent with the assigned structures. Reagents and solvents were purchased from common commercial suppliers and used as received. Column chromatography separations were carried out on Merck silica gel 60 (mesh 230-400). Yields of purified products were not optimized. All starting materials were commercially available unless otherwise indicated.

7-Acetyl-4-butyl-3,4-dihydro-2H-1,4-benzothiazin-3-one tBuOK (0.16 g, 1.45 mmol) was added to a solution of 7-acetyl-3,4dihydro-2H-1,4-benzothiazin-3one^[7] (0.20 g, 0.97 mmol) in dry DMF (3 mL). The mixture was stirred at room temperature for 15 min. then Bul (0.27 q,1.45 mmol) in dry DMF (2 mL) was added dropwise. After being stirred for 4 h, the mixture was poured into ice-chilled water and extracted with EtOAc. The residue

was chromatographed, eluting with cyclohexane/EtOAc 85:15. **1 d** was obtained as oil, (0.18 g, 70 % yield). ^1H NMR (CDCl₃): δ = 0.92 (t, J = 6.9 Hz, 3 H; CH₂CH₃); 1.20–1.45 and 1.50–1.65 (m, each 2 H; CH₂CH₂CH₃); 2.55 (s, 3 H; COCH₃); 3.38 (s, 2 H; SCH₂); 4.01 (t, J = 7.4 Hz, 2 H; NCH₂); 7.17 (d, J = 8.6 Hz, 1 H; H-5); 7.82 (dd, J = 8.6 and 2.1 Hz, 1 H; H-6); 7.95 ppm (d, J = 2.1 Hz, 1 H; H-8). Anal. (C₁₄H₁₇NO₂S) C, H, N.

4-Butyl-7-(1*H***-1-imidazolylacetyl)-3,4-dihydro-2***H***-1,4-benzothiazin-3-one (2 d).** 1) A solution of **1 d** (0.20 g, 0.76 mmol) in CCl₄ (5 mL) was cooled to 0 °C, then Br₂ (0.13 g, 0.84 mmol) in CCl₄ (5 mL) was added dropwise. After being stirred for 1 h at room temperature, the mixture was evaporated to dryness and the residue was chromatographed, eluting with cyclohexane/EtOAc 92:8 to give 7-(bromoacetyl)-4-butyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one as an oil (0.10 g, 40% yield). ¹H NMR (CDCl₃): δ = 0.96 (t, J = 7.2 Hz, 3 H; CH₂CH₃); 1.25-1.50 and 1.55-1.80 (m, each 2 H; CH₂CH₂CH₃); 3.45 (s, 2 H; SCH₂); 3.95-4.15 (m, 2 H; NCH₂); 4.41 (s, 2 H; CH₂Br); 7.15-7.25 (m, 1 H; aromatic H); 7.75-8.05 ppm (m, 2 H; aromatic H). Anal. (C₁₄H₁₆BrNO₂S) C, H, N. 2) Imidazole (0.06 g, 0.88 mmol) was added to a solution of 7-(bromoacetyl)-4-butyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one (0.10 g, 0.29 mmol) in CHCl₃ (3 mL). The mixture was stirred at room temperature for 24 h, then

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evaporated to dryness, and the residue chromatographed eluting with CHCl₃/MeOH 96:4 to give the **2d** (0.82 g, 87% yield) as an orange amorphous solid. $^1\text{H NMR}$ (CDCl₃): $\delta = 0.95$ (t, J = 7.2 Hz, 3 H; CH₂CH₃); 1.20–1.65 and 1.65–1.80 (m, each 2 H; CH₂CH₂CH₃); 3.43 (s, 2 H; SCH₂); 3.95–4.20 (m, 2 H; NCH₂CH₂); 5.40 (s, 2 H; COCH₂N); 6.96, 7.14, and 7.53 (bs, each 1 H; imidazolic H); 7.20–7.30 (m, 1 H; aromatic H); 7.75–8.05 ppm (m, 2 H; aromatic H). Anal. (C₁₇H₁₉N₃O₂S) C, H, N.

4-Butyl-7-{1-[(4-chlorobenzyl)oxy]-2-(1H-1-imidazolyl)ethyl}-4methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one (5 d). 1) NaBH₄ (0.02 g, 0.60 mmol) was added portionwise to a solution of 2d (0.20 g, 0.60 mmol) in dry MeOH (3 mL) over 1 h. The mixture was then evaporated to dryness, and the residue was chromatographed, eluting with CHCl₃/MeOH 92:8 to furnish, 4-butyl-7-[1-hydroxy-2-(1*H*-1-imidazolyl)ethyl]-3,4-dihydro-2*H*-1,4-benzothiazin-3-one as an oil, in quantitative yield. ¹H NMR (CDCl₃): $\delta = 0.95$ (t, J = 7.3 Hz, 3H; CH_2CH_3); 1.30–1.50 and 1.50–1.75 (m, each 2H; $CH_2CH_2CH_3$); 3.37 (s, 2H; SCH₂); 3.80-4.20 (m, 5H; NCH₂CH₂, COCH₂N and CHOH); 4.80-4.95 (m, 1H; CHOH); 6.89 (bs, 2H; imidazolic H); 7.10–7.50 ppm (m, 4H; imidazolic and aromatic H). Anal. (C₁₇H₂₁N₃O₂S) C, H, N. 2) NaH (0.023 g, 0.59 mmol) was added portionwise to a solution of 4-butyl-7-[1-hydroxy-2-(1H-1-imidazolyl)ethyl]-3,4-dihydro-2H-1,4-benzothiazin-3-one (0.13 g, 0.39 mmol) in DMF dry (3 mL) and stirred for 30 min. 4-Chlorobenzyl chloride (0.095 g, 0.59 mmol) in DMF dry (2 mL) was added dropwise. After 90 min the mixture was poured into water and extracted with EtOAc. After evaporation the residue was chromatographed eluting with CH₂Cl₂/MeOH 98:2 to furnish 5 d as an oil (0.12 g, 70 % yield). ¹H NMR (CDCl₃): $\delta = 0.95$ (t, J = 7.4 Hz, 3 H; CH₂CH₃); 1.25–1.50 and 1.50-1.75 (m, each 2H; CH₂CH₂CH₃); 3.37 (s, 2H; SCH₂); 3.80-4.50 ppm (m, 7 H; NCH₂CH₂, CHOCH₂N and OCH₂Ph); 6.75–7.50 (m, 6H; imidazolic and aromatic H). Anal. (C₂₄H₂₆ClN₃O₂S) C, H, N.

General method for the preparation of oxime derivatives 3a–d. The procedure is illustrated by the synthesis of 7-[(1*Z*)-*N*-hydroxy-2-(1*H*-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-3a]. NH₂OH·HCl (0.16 g, 2.30 mmol) was added to a solution of ketone $2a^{[3a]}$ (0.50 g, 1.70 mmol) in EtOH (12 mL), and the pH of the mixture was adjusted to 11 with 15 N NaOH. The solution was refluxed for 3 h, then evaporated to dryness, and the residue was solubilized in H₂O. The aqueous layer was acidified with 1 N HCl to pH 5 and then adjusted to pH 9 with NaHCO₃-saturated aqueous solution. Compound (*Z*)-3a, precipitated out of solution, was crystallized from EtOH (0.27 g, 52%) as a white solid, mp 201–202 °C. ¹H NMR ([D₆]DMSO): δ = 3.38 (s, 3 H; NCH₃); 3.56 (s, 2 H; SCH₂); 5.34 (s, 2 H; CH₂N); 6.88 and 7.12 (s, each 1 H; imidazolic H); 7.30 (d, J=8.6 Hz, 1 H; H-5); 7.55–7.75 (m, 3 H; aromatic and imidazolic H); 12.10 ppm (s, 1 H; NOH). Anal. (C₁₄H₁₄N₄O₂S) C, H, N.

7-[(1*Z*)-*N*-Hydroxy-2-(1*H*-1,2,4-triazol-1-yl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-3 b]. It was synthesized starting from ketone **2** b. [3c] White solid, mp 185–187 °C, 61% yield. ¹H NMR ([D₆]DMSO): δ = 3.29 (s, 3H; NCH₃); 3.45 (s, 2H; SCH₂); 5.45 (s, 2H; CH₂N); 7.27 (d, *J* = 8.7 Hz, 1H; H-5); 7.63 (dd, *J* = 8.7 and 1.3 Hz, 1H; H-6); 7.66 (d, *J* = 1.3 Hz, 1H; H-8); 7.91 and 8.57 (s, each 1H; triazolic H); 12.02 ppm (bs, 1H; NOH). Anal. (C₁₃H₁₃N₅O₂S) C, H, N.

4-Ethyl-7-[(1*Z*)-*N*-hydroxy-2-(1*H*-1-imidazolyl)ethanimidoyl]-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-3 c]. It was synthesized starting from ketone **2 c**. White solid, mp 205.1–205.8 °C, 72% yield. ¹H NMR ([D₆]DMSO): δ = 1.13 (t, J = 6.8 Hz, 3 H; CH₂CH₃); 3.52 (s, 2H; SCH₂); 3.97 (q, J = 6.8 Hz, 2H; CH₂CH₃); 5.32 (s, 2H; NCCH₂N); 6.84 and 7.06 (s, each 1H; imidazolic H); 7.34 (d, J = 8.7 Hz, 1H; H-5); 7.55–7.75 (m, 3 H; aromatic and imidazolic H); 12.12 ppm (s, 1 H; NOH). Anal. (C₁₅H₁₆N₄O₂S) C, H, N.

4-Butyl-7-[(1*Z*)-*N*-hydroxy-2-(1*H*-1-imidazolyl)ethanimidoyl]-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-3 d]. It was synthesized starting from ketone **2 d**. White solid, mp 212–215 °C, 48% yield.

¹H NMR ([D₆]DMSO): δ = 0.86 (t, J=7.2 Hz, 3 H; CH₂CH₃); 1.15–1.60 (m, 4H; CH₂CH₂CH₃); 3.36 (s, 2H; SCH₂); 3.80–4.00 (m, 2H; NCH₂CH₂); 5.31 (s, 2H; NCCH₂N); 6.84 and 7.06 (s, each 1 H; imidazolic H); 7.25–7.75 (m, 4H; aromatic and imidazolic H); 12.10 ppm (s, 1 H; NOH). Anal. (C₁₇H₂₀N₄O₂S) C, H, N.

General method for the preparation of oxime derivatives 3e–g. The procedure is illustrated by the synthesis of 6-[(1*Z*)-*N*-hydroxy-2-(1*H*-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-3 e]. NH₂OH-HCl (0.18 g; 2.60 mmol) was added to a solution of ketone $2e^{|3c|}$ (0.51 g, 1.80 mmol) in pyridine (20 mL). The mixture was stirred at 70 °C for 4 h, evaporated to dryness, and the residue suspended in H₂O. The precipitate was filtered off to afford a crude residue constituted from the mixture of (*Z*)-3e and (*E*)-3e (8.9:1.1) that was chromatographed eluting with CHCl₃ to give the isomer (*Z*)-3e. It was crystallized from MeOH, 0.22 g, 41 % yield, mp 219–220 °C. ¹H NMR ([D₆]DMSO): δ=3.32 (s, 3H; NCH₃); 3.52 (s, 2H; SCH₂); 5.35 (s, 2H; CH₂N); 6.81, 7.04, and 7.65 (s, each 1H; imidazolic H); 7.36–7.42 (m, 3H; aromatic H); 12.10 ppm (s, 1H; NOH). Anal. (C₁₄H₁₄N₄O₂S) C, H, N.

2-Butyl-6-[(1Z)-N-hydroxy-2-(1H-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(Z)-3 f] and 2-butyl-6-[(1E)-N-hydroxy-2-(1H-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(E)-3 f]. They were prepared starting from ketone 2 f. [3d]

(*Z*)-**3 f**, oil, 62.2% yield. 1 H NMR (CDCl₃): δ = 0.88 (t, J = 6.5 Hz, 3 H; CH₂CH₃); 1.25–1.55 (m, 5 H; SCHCHHCH₂CH₂); 1.75–1.95 (m, 1 H; SCHCHHCH₂); 3.25–3.50 (m, 1 H; SCHCH₂); 3.42 (s, 3 H; NCH₃); 5.27 (s, 2 H; CH₂N); 7.02, 7.10, and 7.86 (s, each 1 H; imidazolic H); 7.15–7.37 ppm (m, 3 H; aromatic H). Anal. (C₁₈H₂₂N₄O₂S) C, H, N.

(E)-3 f, oil, 28.7%, 1 H NMR (CDCl₃): δ = 0.86 (t, J = 6.6 Hz, 3 H; CH₂CH₃); 1.20–1.59 (m, 5 H; SCHCHHCH₂CH₂); 1.75–1.95 (m, 1 H; SCHCHHCH₂); 3.34 (s, 3 H; NCH₃); 3.30–3.50 (m, 1 H; SCHCH₂); 4.98 (s, 2 H; CH₂N); 6.89–7.37 (m, 6 H; aromatic and imidazolic H); 12.00 ppm (bs, 1 H; NOH). Anal. (C₁₈H₂₂N₄O₂S) C, H, N.

2-Butoxy-6-[(1Z)-N-hydroxy-2-(1H-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(Z)-3 g] and 2-butoxy-6-[(1E)-N-hydroxy-2-(1H-1-imidazo-lyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(E)-3 g]. They were synthesized starting from ketone 2 g. [3e]

(*Z*)-**3 g**, amorphous solid, 19.2% yield, ¹H NMR (CDCl₃): δ = 0.75 (t, J=6.5 Hz, 3 H; CH₂CH₃); 0.95–1.40 (m, 4 H; CH₂CH₂CH₂CH₃); 3.30–3.55 (m, 1 H; OCHHCH₂); 3.40 (s, 3 H; NCH₃); 3.60–3.80 (m, 1 H; OCHHCH₂); 5.05 (s, 1 H; SCHO); 5.25 (s, 2 H; CH₂N); 7.00–7.45 (m, 5 H; aromatic and imidazolic H); 7.75 (s, 1 H; imidazolic H); 13.75 ppm (bs, 1 H; NOH). Anal. (C₁₈H₂₂N₄O₃S) C, H, N.

(*E*)-**3 g**, amorphous solid, 46.1% yield, ¹H NMR (CDCl₃): δ = 0.75 (t, J=6.5 Hz, 3 H; CH₂CH₃); 1.00–1.50 (m, 4 H; CH₂CH₂CH₂CH₃); 3.30–3.60 (m, 1 H; OCHHCH₂); 3.40 (s, 3 H; NCH₃); 3.65–3.80 (m, 1 H; OCHHCH₂); 5.00 (s, 1 H; SCHO); 5.10 (s, 2 H; CH₂N); 6.90–7.40 (m, 5 H; aromatic and imidazolic H); 7.65 ppm (s, 1 H; imidazolic H). Anal. (C₁₈H₂₂N₄O₃S) C, H, N.

General method for the preparation of oxime ether derivatives 4 a–g. The procedure is illustrated by the synthesis of 7-[(1Z)-N-[(4-chlorobenzyl)oxy]-2-(1H-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(Z)-4a]. NaH (60% mineral oil dispersion, 0.03 g, 0.70 mmol) was added portionwise to a solution of oxime (Z)-3a (0.20 g, 0.66 mmol) in dry DMF (4 mL) under nitrogen atmosphere. The mixture was stirred for 30 min at room temperature, then a solution of 4-chlorobenzyl chloride (0.12 g, 0.66 mmol) in dry DMF (2 mL) was added dropwise. After 20 h the mixture was poured into water and extracted with CHCl₃. The or-

ganic phase was evaporated to dryness and the residue chromatographed eluting with CHCl₃ to give (*Z*)-**4 a** as a yellowish oil, 0.20 g, 71% yield. ¹H NMR (CDCl₃): δ =3.40 (s, 3 H; NCH₃); 3.42 (s, 2 H; SCH₂); 5.18 (s, 2 H; CH₂N); 5.25 (s, 2 H; OCH₂Ph); 6.86, 7.00, and 7.50 (s, each 1 H; imidazolic H); 7.05 (d, J=8.7 Hz, 1 H; H-5); 7.25–7.45 (m, 5 H; aromatic H); 7.60 ppm (d, J=1.8 Hz, 1 H; H-8). Anal. (C₂₁H₁₉CIN₄O₂S) C, H, N.

7-[(1*Z*)-*N*-[(4-Chlorobenzyl)oxy]-2-(1*H*-1,2,4-triazol-1-yl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-4 b]. It was synthesized starting from (*Z*)-3 b. Amorphous solid, 70% yield. ¹H NMR (CDCl₃): δ = 3.43 (s, 3 H; NCH₃); 3.45 (s, 2 H, SCH₂); 5.26 (s, 2 H; CH₂N); 5.39 (s, 2 H; OCH₂Ph); 7.10 (d, *J* = 8.7 Hz, 1 H; H-5); 7.38 (AA′BB′ system, 4H; aromatic H); 7.67 (dd, *J* = 8.7 and 2.1 Hz, 1 H; H-6); 7.82 (d, *J* = 2.1 Hz, 1 H; H-8); 7.93 and 8.11 ppm (s, each 1 H; triazolic H). Anal. (C₂₀H₁₈ClN₅O₂S) C, H, N.

7-[(1*Z*)-*N*-**[(4-Chlorobenzyl)oxy]-2-(1***H*-1-imidazolyl)ethanimidoyl]-4-ethyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-4c]. It was synthesized starting from (*Z*)-3c. Amorphous solid, 80 % yield.

¹H NMR (CDCl₃): δ = 1.26 (t, J = 7.2 Hz, 3 H; CH₂CH₃); 3.38 (s, 2 H; SCH₂); 4.02 (q, J = 7.2 Hz, 2 H; CH₂CH₃); 5.17 (s, 2 H; NCCH₂N); 5.25 (s, 2 H; OCH₂Ph); 6.86, 7.01, and 7.51 (s, each 1 H; imidazolic H); 7.12 (d, J = 7.5 Hz, 1 H; H-5); 7.30–7.45 (m, 5 H; aromatic H); 7.61 ppm (d, J = 1.4 Hz, 1 H; H-8). Anal. (C₂₂H₂₁ClN₄O₂S) C, H, N.

4-Butyl-7-[(1*Z)-N-*[(**4-chlorobenzyl)oxy]-2-(1***H*-1-imidazolyl)ethanimidoyl]-**3,4-dihydro-2***H*-**1,4-benzothiazin-3-one** [(*Z*)-**4d**]. It was synthesized starting from (*Z*)-**3 d**. Amorphous solid, 65% yield.

¹H NMR (CDCl₃): δ = 0.94 (t, J = 7.3 Hz, 3 H; CH₂CH₃); 1.20–1.45 (m, 2 H; CH₂CH₂CH₃); 1.50–1.65 (m, 2 H; CH₂CH₂CH₃); 3.39 (s, 2 H; SCH₂); 3.99 (t, J = 7.7 Hz, 2 H; NCH₂CH₂); 5.18 (s, 2 H; OCH₂Ph); 5.27 (s, 2 H; NCCH₂N); 6.88, 7.03, and 7.52 (s, each 1 H; imidazolic H); 7.10 (d, J = 8.7 Hz, 1 H; H-5); 7.30–7.45 (m, 5 H; aromatic H); 7.62 ppm (d, J = 2.1 Hz, 1 H; H-8). Anal. (C₂₄H₂₅CIN₄O₂S) C, H, N.

6-[(1Z)-N-[(4-Chlorobenzyl)oxy]-2-(1H-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2H-1,4-benzothiazin-3-one [(Z)-4 e]. It was synthesized starting from (Z)-3 e. Oil, 58 % yield. 1 H NMR (CDCl₃): δ = 3.39 (s, 5 H; NCH₃ and SCH₂); 5.19 (s, 2 H; CH₂N); 5.25 (s, 2 H; OCH₂Ph); 6.84, 7.00, and 7.50 (s, each 1 H; imidazolic H); 7.14 (dd, J=8.0 and 1.8 Hz, 1 H; H-6); 7.23 (d, J=1.8 Hz, 1 H; H-8); 7.25–7.40 ppm (m, 5 H; aromatic H). Anal. ($C_{21}H_{19}\text{CIN}_4O_2\text{S}$) C, H, N.

2-Butyl-6-[(1*Z*)-*N*-**[(4-chlorobenzyl)oxy]-2-(1***H*-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-4 f]. It was synthesized starting from (*Z*)-3 f. Oil, 47.6% yield.

¹H NMR (CDCl₃): δ = 0.87 (t, J = 7.3 Hz, 3 H; CH₂CH₃); 1.26–1.50 (m, 5 H; SCHCHHCH₂CH₂); 1.80–1.95 (m, 1 H; SCHCHHCH₂); 3.30–3.50 (m, 1 H; SCHCH₂); 3.42 (s, 3 H; NCH₃); 5.24 (s, 2 H; CH₂N); 5.27 (s, 2 H; OCH₂Ph); 6.83, 7.05, and 7.74 (s, each 1 H; imidazolic H); 7.14–7.40 ppm (m, 7 H; aromatic H). Anal. (C₂₅H₂₇ClN₄O₂S) C, H, N.

2-Butyl-6-[(1*E***)-***N***-[(4-chlorobenzyl)oxy]-2-(1***H***-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2***H***-1,4-benzothiazin-3-one [(***E***)-4 f]. It was synthesized starting from (***E***)-3 f. Oil, 50 % yield. ¹H NMR (CDCl₃): \delta = 0.87 (t, J = 7.3 Hz, 3 H; CH₂CH₃); 1.25–1.55 (m, 5 H; SCHCHHCH₂CH₂); 1.75–1.95 (m, 1 H; SCHCHHCH₂); 3.30–3.50 (m, 1 H; SCHCH₂); 3.42 (s, 3 H; NCH₃); 5.12 (s, 2 H; CH₂N); 5.27 (s, 2 H; OCH₂Ph); 6.90–7.79 ppm (m, 10 H; aromatic and imidazolic H). Anal. (C₂₅H₂₇ClN₄O₂S) C, H, N.**

2-Butoxy-6-[(1*Z*)-*N*-[(4-chlorobenzyl)oxy]-2-(1*H*-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2*H*-1,4-benzothiazin-3-one [(*Z*)-4g]. It was synthesized starting from (*Z*)-3g. Oil, 91% yield. 1 H NMR (CDCl₃): δ =0.77 (t, J=7.0 Hz, 3 H; CH₂CH₃); 1.05–1.50 (m, 4H; CH₂CH₂CH₂CH₃); 3.35–3.46 (m, 1H; OCHHCH₂); 3.46 (s, 3 H; NCH₃); 3.60–3.70 (m, 1 H; OCHHCH₂); 5.05 (s, 1 H; SCHO); 5.19 (s, 2 H; CH₂N); 5.26 (s, 2 H; OCH₂Ph); 6.85, 7.01, and 7.56 (s, each 1 H; imidazolic H); 7.18–7.39 ppm (m, 7 H; aromatic H). Anal. (C_{25} H₂₇ClN₄O₃S) C, H, N.

2-Butoxy-6-[(1*E***)-***N***-[(4-chlorobenzyl)oxy]-2-(1***H***-1-imidazolyl)ethanimidoyl]-4-methyl-3,4-dihydro-2***H***-1,4-benzothiazin-3-one [(***E***)-4g]. It was synthesized starting from (***E***)-3g. Oil, 63% yield.

¹H NMR (CDCl₃): \delta = 0.77 (t, J = 7.4 Hz, 3 H; CH₂CH₃); 1.06–1.45 (m, 4H; CH₂CH₂CH₂CH₃); 3.35–3.55 (m, 1H; OCHHCH₂); 3.46 (s, 3 H; NCH₃); 3.65–3.75 (m, 1H; OCHHCH₂); 5.06 (s, 1H; SCHO); 5.21 (s, 2H; CH₂N); 5.26 (s, 2H; OCH₂Ph); 6.86, 7.02, and 7.62 (s, each 1 H; imidazolic H); 7.10–7.45 ppm (m, 7 H; aromatic H). Anal. (C_{25}H_{27}CIN_4O_3S) C, H, N.**

Computational Method

A chimeric model of the *C. albicans* CYP51 was constructed following the computational procedure described in Rossello et al.^[6e] The resulting structure of the enzyme was prepared for docking experiments using the protein preparation wizard script as implemented in Maestro 7.5.^[10] Docking experiments were carried out using the default settings of Glide 4.0^[11] and the standard precision procedure. The five best poses for each compound were retrieved from an overall search of 50 000 possible poses. The search was performed within a grid cage the size of which was calculated in proportion to the crystal structure of CYP51 in complex with fluconazole. During docking experiments, a search constraint was inserted on the iron atom of the heme group.

Susceptibility testing

Susceptibility testing was performed by the M27-A micro-dilution method of the National Committee for Clinical Laboratory Standards $^{[12]}$ in 0.165 $\,\mathrm{M}$ MOPS (morpholinepropanesulfonic acid)-buffered (pH 7) RPMI 1640 medium (Gibco BRL, Paisley, United Kingdom). The activity of compounds against *C. albicans* and *C. krusei* was tested using serial dilutions ranging from 0.9 to 500 $\,\mathrm{\mu g}\,\mathrm{mL}^{-1}$. The MIC was the lowest concentration of chemical that produced an 80% reduction in the turbidity compared to chemical-free normal subjects.

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